

Law Offices
FOLEY & LARDNER
Suite 500
3000 K Street, N.W.
Washington, DC 20007-5109
(202) 672-5300

jc503 U.S. PTO
09/344382
06/25/99

TO: Assistant Commissioner for Patents
Box Patent Applications
Washington D.C. 20231

06/25/99
U.S. PTO

Attorney Docket No. 050499/0101
(must include alphanumeric codes if no inventors named)

UTILITY PATENT APPLICATION TRANSMITTAL
(new nonprovisional applications under 37 CFR 1.53(b))

Transmitted herewith for filing is the patent application of:

**INVENTOR(S): Shunichi SOMA; Masahiro IWAMOTO; Kojiro KURISU; and
Yoshinobu HIGUCHI**

TITLE: ORTHODONTIC REMEDIES CONTAINNG PTH

In connection with this application, the following are enclosed:

APPLICATION ELEMENTS:

XX Specification - 27 TOTAL PAGES

(preferred arrangement:)

- Descriptive Title of the Invention
- Cross Reference to Related Applications
- Statement Regard Fed sponsored R&D
- Reference to Microfiche Appendix
- Background of the Invention
- Brief Summary of the Invention
- Brief Description of the Drawings (if filed)
- Detailed Description
- Claim(s)
- Abstract of the Disclosure

XX Drawings - Total Sheets 18

XX Declaration and Power of Attorney - Total Sheets 4

XX Newly executed (original or copy)

Copy from a prior application (37 CFR 1.63(d))

(relates to continuation/divisional boxes completed) - NOTE: Box below

DELETION OF INVENTOR(S) - Signed statement attached deleting inventor(s)
named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b).

Incorporation By Reference (useable if copy of prior application
Declaration being submitted)

The entire disclosure of the prior application, from which a COPY of the
oath or declaration is supplied as noted above, is considered as being
part of the disclosure of the accompanying application and is hereby
incorporated by reference therein.

Microfiche Computer Program (Appendix)

Nucleotide and/or Amino Acid Sequence Submission (if applicable,
all necessary)

Computer Readable Copy

Paper Copy (identical to computer copy)

Statement verifying identify of above copies

ACCOMPANYING APPLICATION PARTS

- Assignment Papers (cover sheet & document(s))
 37 CFR 3.73(b) Statement (when there is an assignee)
 English Translation Document (if applicable)
 Information Disclosure Statement (IDS) with PTO-1449. _____ Copies of IDS Citations
 Preliminary Amendment
 Return Receipt Postcard (MPEP 503)
 Small Entity Statement(s)
_____ Statement file in prior application, status still proper and desired.
 Certified Copy of Priority Document(s) with Claim of Priority
_____ (if foreign priority is claimed).
 OTHER: Check in the amount of \$818.00

If a **CONTINUING APPLICATION**, check appropriate box and supply the requisite information:

Continuation Divisional Continuation-in-part (CIP)
of prior application Serial No. ____.

Amend the specification by inserting before the first line the following sentence: --This application is a _____ continuation, _____ divisional or _____ continuation-in-part of application Serial No. ____,
filed ____.--

CORRESPONDENCE ADDRESS:

Foley & Lardner Address noted above.
Telephone: (202) 672-5300
Fax Number: (202) 672-5399

FEE CALCULATIONS: (Small entity fees indicated in parentheses.)

(1) For	(2) Number Filed	(3) Number Extra	(4) Rate	(5) Basic Fee \$760 (\$380)
Total Claims	21 - 20 =	1	x \$18 (x \$9)	\$18.00
Independent Claims	3 - 3 =	0	x \$78 (x \$39)	
Assignment Recording Fee per property			\$40	\$40.00
Surcharge Under 37 C.F.R. 1.16(e)			\$130 (\$65)	
				TOTAL FEE: \$818.00

METHOD OF PAYMENT:

A check in the amount of the above TOTAL FEE is attached. If payment is enclosed, this amount is believed to be correct; however, the Commissioner is hereby authorized to charge any deficiency or credit any overpayment to Deposit Account No. 19-0741.

Respectfully submitted,


Stephen A. Bent
Reg. No. 29,768

Date: June 25, 1999
Docket No.: 050499/0101

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No. 050499/0101

In re patent application of

Shunichi SOMA et al.

Serial No. UNASSIGNED

Filed: CONCURRENTLY HEREWITH

For: ORTHODONTIC REMEDIES CONTAINING PTH

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to examination of the above-identified application, Applicants respectfully request that the following amendments be entered into the application:

IN THE SPECIFICATION

On page 4, between lines 14 and 15, please insert the following paragraph:

-- The present invention further relates to a kit comprising an effective amount of a parathyroid hormone or a derivative thereof and instructions for increasing tooth movement. --

IN THE CLAIMS

Please cancel claims 1-21 without prejudice or disclaimer.

Please add the following new claims:

-- 22. A method for increasing tooth movement comprising administering to a subject in need thereof an effective amount of a parathyroid hormone or a derivative thereof.

23. The method as claimed in claim 22, wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 84.

24. The method as claimed in claim 23, wherein the parathyroid hormone is substantially pure parathyroid hormone.
25. The method as claimed in claim 23, wherein the parathyroid hormone is recombinant parathyroid hormone.
26. The method as claimed in claim 22, wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 34.
27. The method as claimed in claim 26, wherein the parathyroid hormone is substantially pure parathyroid hormone.
28. The method as claimed in claim 26, wherein the parathyroid hormone is recombinant parathyroid hormone.
29. A kit for increasing tooth movement comprising an effective amount of a parathyroid hormone or a derivative thereof and instructions for increasing tooth movement.
30. The kit as claimed in claim 29, wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 84.
31. The kit as claimed in claim 30, wherein the parathyroid hormone is substantially pure human parathyroid hormone.
32. The kit as claimed in claim 30, wherein the parathyroid hormone is recombinant parathyroid hormone.
33. The kit as claimed in claim 29, wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 34.
34. The kit as claimed in claim 33, wherein the parathyroid hormone is substantially pure parathyroid hormone.
35. The kit as claimed in claim 33, wherein the parathyroid hormone is recombinant parathyroid hormone.
36. In an orthodontic composition, the improvement comprising an effective amount of a parathyroid hormone or a derivative thereof for increasing tooth movement.
37. The orthodontic composition as claimed in claim 36, wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 84.

38. The orthodontic composition as claimed in claim 37, wherein the parathyroid hormone is substantially pure parathyroid hormone.

39. The orthodontic composition as claimed in claim 37, wherein the parathyroid hormone is recombinant parathyroid hormone.

40. The orthodontic composition as claimed in claim 36, wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 34.

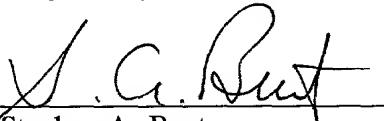
41. The orthodontic composition as claimed in claim 40, wherein the parathyroid hormone is substantially pure parathyroid hormone.

42. The orthodontic composition as claimed in claim 40, wherein the parathyroid hormone is recombinant parathyroid hormone.—

REMARKS

The Examiner is respectfully requested to enter the above amendments prior to examination of the instant application.

Respectfully submitted,


Stephen A. Bent
Reg. No. 29,768

June 25, 1999
Date

FOLEY & LARDNER
3000 K Street, N.W.
Suite 500
Washington, D.C. 20007-5109
Tel: (202) 672-5300

ORTHODONTIC REMEDIES CONTAINING PTH

TECHNICAL FIELD

This invention relates orthodontic remedies containing
5 parathyroid hormone (PTH) or PTH derivatives as the active
ingredient.

BACKGROUND ART

Parathyroid hormone (PTH) is known as one of the hormones which plays an important role in bone metabolism.
10 There have been reported a number of effects of PTH on bones. In the field of clinical orthodontics, tooth movement is considered to be an accelerated remodeling of a bone due to mechanical stress acting on the tooth. The adaptation of an alveolar bone to such mechanical stress has been shown to
15 constitute an increase in bone resorption in the pressured side of the periodontia and an increase in the bone formation in the strained side of the periodontia. Although attempts have been made to give a clear explanation of these changes taking place in the periodontia under mechanical
20 stress on the basis of the tension hypothesis (Oppenheim, 1911), a detailed cell response mechanism to mechanical stress has not yet been clarified so far (Sandy, Farndale and Meikle, 1993).

It has been considered that acceleration of the bone
25 turnover at a tooth movement is an important factor relating to orthodontic tooth movement. This is because the treatment period can be shortened by accelerating bone turnover. It has been reported that bone resorption can be

accelerated in experimental tooth movement by locally or topically administering chemicals such as PGE₁ (Yamasaki, Miura and Suda, 1980; Lee, 1990), PGE₂ (Yamasaki, Miura and Suda, 1980; Chao et al., 1988) and 1 α ,25-(OH)₂D₃ (Collins and Sinclair, 1988; Takano-Yamamoto, et al., 1992) or systemically administering PGE₁ (Lee, et al., 1988).

It is well known that parathyroid hormone (PTH) is one of the systemic factors required in bone remodeling. Intermittent injection of PTH in vivo brings about an increase in bone mass of ovariectomized (OVX) rats (Hock, et al., 1988; Liu, et al., 1991; Ibbotson, 1992) or normal rats (Hock and Gera, 1992; Dobnig, 1995). It is, therefore, considered that stimulation of bone formation is one of the physiological roles of the pulsating secretion of PTH in vivo. On the contrary, the results of morphological bone measurement indicate that continuous injection of PTH results in the simultaneous acceleration of bone formation and bone resorption but no substantial increase in bone mass either in parathyroidectomized rats (Kitagawa, et al., 1991) and normal dogs (Malluche, et al., 1982). According to the data obtained from various studies in vivo, osteoclast formation (Takahashi, et al., 1988; Kurihara, et al., 1991) and osteoblast proliferation (Somjen, et. al, 1990) are both stimulated by PTH. It is also reported that the administration of PTH exerts systemic effects on the periodontia and, therefore, causes different findings in the alkaline phosphatase reaction in the periodontal ligaments and osteoclast distribution compared with a control group

(T.Deguchi, J. Japan Orthodontic Dentistry, Vol. 28, No. 1, 1969, pp. 1-7).

With respect to the role of PTH in bone remodeling relating to orthodontic tooth movement, it has been proved
5 in the above-cited report (Kamata, 1972) that the induction of osteoclasts in the pressured side during an experimental tooth movement is completely inhibited by para-thyroidectomy and then restored by injecting a parathyroid extract. This fact indicates that PTH would play an important role in the
10 osteoclast formation during the experimental tooth movement. However, practical application of PTH in the field of clinical orthodontics has never been clarified hitherto.

DISCLOSURE OF THE INVENTION

An object of the present invention is to provide
15 orthodontic remedies which are practically usable and efficacious in the field of clinical orthodontics.

After conducting extensive studies, the present inventors have found that orthodontic tooth movement is accelerated by administering parathyroid hormone (PTH) or
20 one or more PTH derivatives, thus completing the present invention. Accordingly, the present invention relates to orthodontic remedies containing parathyroid hormone (PTH) or one or more PTH derivatives as the active ingredient(s). The present invention further relates orthodontic remedies containing human PTH (1-84) or one or more derivatives thereof as the active ingredient(s). The present invention further relates to orthodontic remedies containing human PTH (1-34) or one or more derivatives thereof as the active

ingredient(s). Furthermore, the present invention relates orthodontic remedies characterized by containing parathyroid hormone (PTH) as the active ingredient. The present invention further relates to orthodontic remedies containing 5 human PTH (1-84) as the active ingredient. The present invention further relates to orthodontic remedies containing human PTH (1-34) thereof as the active ingredient. In addition, the present invention relates to dental compositions containing parathyroid hormone (PTH) or one or 10 more PTH derivatives as the active ingredient(s). Further, the present invention relates to noninvasive PTH preparations characterized by the continuos administration of parathyroid hormone (PTH) or one or more PTH derivatives in an efficacious amount.

15 The terms "orthodontic dentistry" and "orthodontics" are used herein interchangeably.

The term "orthodontic remedy" as used herein means a drug to be used for correcting abnormalities in teeth or upper and/or lower jaws. The orthodontic remedies of the 20 present invention are employed preferably as drugs for correcting dental irregularities, i.e., remedies for dental irregularity. The term "remedy for dental irregularity" as used herein means a drug to be used for shifting a specific tooth with an abnormality or all teeth into the normal position to thereby normalize a dental arch suffering from some morphological abnormality (i.e., dental irregularity), for example, abnormal interdental distance, tooth malposition (dislocation toward lip (cheek) or tongue).

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a diagram which shows a method for experimentally shifting a tooth by using an elastic band.

Fig. 1 (A) shows a method wherein the elastic band is inserted between the first molar (M_1) and the second molar (M_2) by the Wong and Rothblatt method, while (B) and (C) show a method for measuring the distance between M_1 and M_2 with the use of a contact gauge.

Fig. 2 is a graph which shows a dose-dependent effect of PTH (1-84) infusion on teeth separation.

Fig. 3 is a graph which shows a change with the passage of time in the effect of 10 $\mu\text{g}/100 \text{ g/day}$ of PTH (1-84) infusion into rats on teeth separation.

Fig. 4 is a graph which shows a dose-dependent effect of PTH (1-84) infusion on the appearance of osteoclasts in teeth separation.

Fig. 5 is a graph which shows a change over time in the effect of 10 $\mu\text{g}/100 \text{ g/day}$ of PTH (1-84) infusion on the appearance of osteoclasts in the pressured side, when an elastic band is inserted between the first molar and the second molar.

Fig. 6 is a diagram which shows a method for orthodontic tooth movement with the use of a closed coil spring. Fig. 6 (A) and (B) show a method wherein an ultra-elastic closed coil spring is ligated between the upper incisive tooth and the right first molar for traction, while (C) shows a method for measuring the shift distance of the tooth with the use of a calipers under a stereoscopic

microscope.

Fig. 7 is a graph which shows a dose-dependent effect of hPTH (1-34) infusion on the mesial shift of the first molar.

5 Figs. 8 and 9 are photographs which show effects of continuous hPTH (1-34) infusion and intermittent hPTH (1-34) injection on the mesial shift of the first molar.

10 Fig. 10 is a graph which shows changes over time in the effects of continuous hPTH (1-34) infusion and
10 intermittent hPTH (1-34) injection on the mesial shift of the first molar.

Fig. 11 is a graph which shows an effect of local injection of sustained release hPTH (1-34) on the mesial shift of the first molar.

15 Figs. 12 to 14 are photographs which show effects of local injection of hPTH (1-34) on the shift of the first molar.

20 Fig. 15 is a graph which shows a change with the passage of time in the effect of local injection of hPTH (1-34) on the shift of the first molar.

Figs. 16 and 17 are photographs which show histological findings of the effects of systemic continuous infusion of hPTH (1-34) and intermittent injection of hPTH (1-34) on the shift of the first molar.

25 Fig. 18 is a photograph which shows histological findings of the effect of local injection of hPTH (1-34) on the shift of the first molar.

BEST MODE FOR CARRYING OUT THE INVENTION

The parathyroid hormone (PTH) to be used in the present invention involves natural PTH, recombinant PTHs produced by genetic engineering techniques and chemically synthesized PTHs. Preferable examples thereof include human PTH consisting of 84 amino acid residues (human PTH (1-84)), in particular, recombinant human PTH (1-84) produced by genetic engineering techniques. The term "PTH derivative" involve peptide fragments of the above-mentioned PTHs; peptides constructed by partly substituting the amino acids constituting PTH per se or a peptide fragments thereof by other amino acids; those constructed by partly deleting the amino acids constituting PTH per se or a peptide fragments thereof; and those constructed by adding one or more amino acids to PTH per se or a peptide fragments thereof, each having the same activity. Examples of the peptide fragments of PTH include human PTH (1-34), human PTH (1-64), human PTH (35-84) and bovine PTH (1-34). PTH (1-34) means a peptide fragment of PTH having an amino acid sequence ranging from the amino acid at the N-terminus to the one at the 34-position. As a preferable example of the peptide fragments of PTH, human PTH consisting of 34 amino acid residues (i.e., human PTH (1-34)), in particular, a recombinant human PTH (1-34) constructed by genetic engineering techniques may be cited.

Preferable examples of the amino acid substitution include the substitution of the constituting amino acid at the 8-position by leucine or norleucine, the substitution of

the constituting amino acid at the 18-position by leucine or norleucine, and the substitution of the constituting amino acid at the 34-position by tyrosine.

Preferable examples of the parathyroid hormone (PTH) 5 or PTH derivatives to be used in the orthodontic remedies, dental compositions or noninvasive PTH preparations according to the present invention include human PTH (1-84), human PTH (1-34), human PTH (1-38), human PTH (1-37) and human PTH (1-34)-NH₂. Among all, human PTH (1-84) and human 10 PTH (1-34) are still preferable and human PTH (1-34) may be cited as the most desirable one.

It is not always necessary for the parathyroid hormone (PTH) or PTH derivatives to be used in orthodontic remedies, dental compositions or noninvasive PTH preparations 15 according to the present invention have a purity of 100%. Namely, these PTH and PTH derivatives may be substantially pure ones. The term "substantially pure" as used herein means having been purified at least to such an extent as showing a single peak in HPLC, preferably having been 20 identified as being uniform by combining procedures such as SDS-DAGE, capillary electrophoresis, etc. Such PTHs can be proved and identified also by using a method disclosed in JP(Kokai) Hei-6-87897 or methods described in Domestic Announcement No. 4-505259 and J. Biol. Chem., 265, 15854 25 (1990) which are optionally modified.

The drugs of the present invention may have a dosage form of injections (solutions, freeze-dried preparations, etc.) obtained by a method conventionally employed in

producing peptide preparations. Alternatively, they may be
in the form of preparations with localized and delayed
actions, for example, oral transmucosal preparations
produced by packing the drugs in microcapsules or
5 impregnating gel sheets therewith. In the preparation, use
can be made of pharmaceutically acceptable auxiliary
ingredients. It is also possible to modify the preparations
with polyethylene glycol so as to prolong the half-life in
blood. Preferable examples of the preparations are
10 noninvasive ones.

Oral transmucosal preparations have been put into
practical use with respect to nitroglycerin, nicotin,
nifedipine, etc. The advantages of such oral transmucosal
preparations as noninvasive peptide or protein preparations
15 reside in that they can be conveniently administered without
resort to any specific device, that they suffer from neither
digestion in the digestive tract nor the first pass effect
in the liver, etc. In the case of hydrophilic substances
such as peptides and proteins, however, it is needed to use
20 enhancers to pass through the physicochemical and enzymatic
barriers in the oral mucosa, different from the low-
molecular weight compounds as cited above. As the
enhancers, use can be made of bile acids, dihydrofusidic
acids, cyclodextrins, surfactants and chelating agents.

25 Examples of auxiliary ingredients usable in the
preparations of the present invention include bases,
stabilizers, antiseptics, preservatives, emulsifiers,
suspending agents, solubilizing agents, solubilizing aids,

lubricating agents, corrigents, colorants , perfumes, soothing agents, vehicles, binders, thickening agents and buffer agents. More particularly speaking, it is possible therefor to use, for example, calcium carbonate, lactose, 5 sucrose, sorbitol, mannitol, starch, amylopectin, cellulose derivatives, gelatin, cacao fat, distilled water for injection, aqueous solution of sodium chloride, Ringer's solution, glucose solution, human serum albumin, etc.

To produce the drugs according to the present 10 invention with the use of these auxiliary ingredients, appropriate ones may be selected from among these auxiliary ingredients and employed as stated in, for example, "Iyakuhin Tenkabutsu Ichiran-hyo (List of Pharmaceutical Additives)" (published by Zaidan Hojin Tokyo Iyakuhin Kogyo Kyokai Iji Hoki Iinkai (Committee of Legal Provisions on Medical Affairs, Foundation of Tokyo Pharmaceutical Industry Association) & Osaka Iyakuhin Kogyo Kyokai Iji Hoki Kenkyu Iinkai (Committee of Legal Provisions on Medical Affairs, 15 Osaka Pharmaceutical Industry Association). The amount of each auxiliary ingredient may be appropriately determined 20 within the pharmaceutically acceptable range depending on the dosage form, etc.

The drugs of the present invention may be administered either locally or topically or systemically. When a 25 definite tooth (for example, a front tooth is to be exclusively shifted, local or topical administration is preferable. In particular, continuous local or topical administration is preferable therefor. Preferable examples

of methods for the continuous local or topical administration involve topical infusion of PTH with the use of sustained release bases or continuous transmucosal absorption of PTH. Particular examples of the sustained

5 release bases include those usable in submucosal or subperiosteal infusion such as (1) collagen pellets, (2) polylactic acid bases, (3) hydroxyapatite cement, and (4) alginic acid gel. Alternatively, use may be made of patches for transmucosal administration. On the other hand, the

10 advantage of the systemic administration resides in that PTH can be administered without any invasion by using an elaborately planned administration method of this type. Preferable examples of the systemic administration method include subcutaneous administration, intravenous

15 administration, nasal administration and transpulmonary administration. When it is desired to quickly shift all teeth, systemic administration is seemingly superior in convenience to local or topical administration over a broad scope.

20 The administration period may be determined depending on the cause of the diseases by a clinical dentist based on the period required for shifting the target tooth and fixing the thus shifted alveolar bone. The administration frequency may range from once three months to everyday. It

25 is preferable to administer PTH once a month to 5 times per week, or everyday. Continuous administration is particularly preferable.

The administration dose of PTH according to the

present invention may vary depending on the tooth shifting distance, tooth type, the number of the teeth to be shifted, etc. In the case of systemic administration, the dose of PTH ranges from 0.1 µg to about 10 mg, preferably from 10 µg
5 to 1 mg.

Examples

To further illustrate the present invention in greater detail, the following Examples will be given. However, it is to be understood that the present invention is not
10 restricted thereto.

Materials

Details (type and manufacturer) of the animals and chemicals employed in these examples were as follows. Male Wister rats (350 to 400 g) were obtained from Oriental Yeast Co., Ltd. (Tokyo). PTH employed in Example 1 was recombinant human PTH (1-84) which had been produced by using a modification of the methods described in Domestic Announcement No. 4-505259 and J. Biol. Chem., 265, 15854 (1990). PTH employed in Example 2 was recombinant human PTH (1-34) manufactured by Peptide Institute Inc., (Mino).
15 Osmotic pumps (Alzet 2ML1) (were purchased from Alza (Palo Alto, Calif. USA). Tween-80 was purchased from Wako Pure Chemical Industries, Ltd. (Tokyo). Elastic bands for orthodontics (Quick-Stik, A-1) were purchased from Unitek
20 (Monrovia, Calif., USA).
25

Method of experimental tooth movement

There are two methods for experimental tooth movement, i.e., one wherein an elastic band (made of rubber) is

inserted between teeth and another one wherein a closed coil spring for orthodontics is used. The closed coil spring employed herein is Sentalloy closed coil spring 509-21 (made of Ni-Ti, manufactured by Tomy International K.K.) which can give an almost constant traction force of about 30 g within a spring-elongation range of 2 to 3 mm. That is to say, it shows no increase in the traction force in proportion to the spring-elongation, as observed in conventional springs.

When the initial elongation of the spring is regulated within the above-mentioned range, therefore, a constant force can be applied regardless of orthodontic tooth movement. When a rubber substance is inserted between molars, an extremely large force is applied immediately after the insertion but scarcely any force is applied after the target tooth is shifted by 0.5 mm. Accordingly, either the method with the insertion of an elastic (rubber) band or another one with the use of a closed coil spring may be used in experimental tooth movement within a short period of time. When a tooth is to be shifted over a long time, however, it is appropriate to use the latter method wherein the first molar is mesially shifted by elongating a closed coil spring. The method with the insertion of an elastic (rubber) band was employed in Example 1, while another method with the use of a closed coil spring was employed in Example 2.

25 Example 1

Experiment 1: Dose-dependent effects of PTH infusion on experimental tooth movement

18 rats were divided into 4 groups including a control

group having 6 animals and 3 PTH-infusion groups each having 4 animals. PTH was dissolved in a citric acid-buffered saline containing 0.05% of Tween 80 and introduced into osmotic pumps. Then these pumps were implanted in the 5 subcutus posterior region of the neck of the rats under ether anethesia. PTH was continuously infused into the rats in doses of 1, 3 and 10 $\mu\text{g}/100\text{ g}$ body weight/day and the rats were fed with a standard pelletized feed (manufactured by Oriental Yeast Co., Ltd.). To the rats of the control 10 group, vehicles were exclusively administered. 48 hours after implanting, a piece of an elastic band (0.8 mm in thickness) was inserted between the right upper first molar and second molar (between M_1 and M_2) of each rat under ether anethesia in accordance with the method of Wong-Rothblatt 15 (1954) as shown in Fig. 1(A). On day 3 of the teeth separation, the rat was sacrificed by ether-inhalation. After cutting out the upper jaw, the distance between the adjacent faces of M_1 and M_2 was measured with the combined use of contact gauges (manufactured by Sun Dental, Osaka) of 20 50, 100 and 150 μm in thickness (Fig. 1 (B) and (C)). In the case of the control group, a contact gauge of 50 μm could be inserted between M_1 and M_2 . Thus, the interdental distance was calculated by subtracting 50 μm from the measured 25 distance of each animal. Fig. 2 shows the results wherein "*" means that a significant difference from the control group (PTH administration: 0 $\mu\text{g}/100\text{ g}$ body weight/day) is observed at a significance level of 5%. Fig. 2 indicates that the maximum effect was achieved by infusing PTH in a

dose of 10 µg/100 g body weight/day. The upper jaw was fixed in 4% paraformaldehyde, decalcified in 4% formic acid and then implanted in paraffin. Subsequently, these preparations were cut into continuous mesiodistal sections 5 of 8 µm in thickness and stained with hematoxylin and eosin. Histological examinations were all performed in the pressured side of the M₁ interradicular septa and osteoclasts in the area of 300 x 700 µm² in this region were counted (Fig. 1 (C)). Osteoclasts were counted based on the 10 fact that they were large multinuclear cells having been stained with eosin and located adjacent to the bone surface. The statistic differences between the control group and the test group was evaluated in accordance with Wilcoxon's rank-sum calibration method. Each of the data was expressed in 15 average ± SEM. A P value less than 0.05 was regarded as being statistically significant.

Experiment 2: Change with the passage of time in effects of PTH infusion on experimental tooth movement

32 rats were divided into 2 test groups each having 16 animals. PTH was infused into these rats in a dose of 10 µg/100 g body weight/day, since it had been revealed by the 20 above Experiment 1 that the maximum effect could be established at this dose. To the rats of the control group, vehicles were exclusively administered. 2 days after 25 implanting osmotic pumps, an elastic band was inserted between the right M₁ and M₂ of each rat. On days 0, 1, 3 and 5 of the teeth separation (i.e., on days 2, 3, 5 and 7 of the PTH infusion), the rats were sacrificed followed by the

same procedures as those performed in Experiment 1.

Results

1. Effects of PTH (1-84) infusion on teeth separation due to elastic band:

5 Fig. 2 shows the dose-dependent effect of PTH (1-84) infusion on teeth separation. As reported in a number of papers, teeth separation between M_1 and M_2 arose after inserting the elastic band for 3 days. Compared with the rats of the control group, the rats to which 10 $\mu\text{g}/100\text{ g}$ body weight/day of PTH was infused showed a significant increase in the distance between M_1 and M_2 . Fig. 3 shows a change with the passage of time in the effect of 10 $\mu\text{g}/100\text{ g}$ body weight/day of PTH (1-84) infusion into rats on teeth separation wherein "*" means that a significant difference 10 from the control group on day 3 was observed at a significance level of 5%. On day 1, no significant difference was observed between the rats of the control group and the PTH-treated rats. On day 3, however, the treated rats showed a significant increase in the separation 15 20 25

distance. On day 5 of the teeth separation, the separation in the PTH-treated rats seemingly almost reached the limit. On day 5, scarcely any friction was observed between the teeth and the elastic band in the PTH-treated rats and, therefore, the experiment was ceased.

2. Effect of continuous PTH (1-84) infusion on the number of osteoclasts in the pressured side of periodontia:

Fig. 4 shows a dose-dependent effect of PTH (1-84) infusion on the appearance of osteoclasts in teeth

separation wherein "*" means that a significant difference from the control group (PTH administration: 0 µg/100 g body weight/day) is observed at a significance level of 5%. As having been reported in a number of papers, osteoclasts in
5 the pressured side of the periodontia were increased after inserting the elastic band for 3 days. Different from the results with respect to the teeth separation, the control group showed a significant increase in the osteoclast count compared with all of the 3 test groups.

10 Fig. 5 shows a change with the passage of time in the effect of 10 µg/100 g body weight/day of PTH (1-84) infusion on the appearance of osteoclasts in the pressured side, wherein "*" means that a significant difference from the control group was observed on each day at a significance
15 level of 5%. The osteoclast count showed a significant increase from day 1 of the PTH infusion. On day 5, no significant difference was observed in the osteoclast count between the rats of the control group and the PTH-treated ones.

20 3. Histological change in pressured periodontia of rats with continuous PTH (1-84) infusion:

Histological remodelings of the pressured periodontia in the teeth separation were located exclusively in the mesial periodontium of the mesiobuccal face of M_1 . On day 1
25 of the teeth separation, necrotic tissue was observed both in the vehicle-treated rats (i.e., the rats of the control group) and the rats treated with 10 µg/100 g body weight/day of PTH. On day 3 of the teeth separation, the vehicle-

treated rats still showed necrotic tissue in the pressured side. However, the rats treated with 10 µg/100 g body weight/day of PTH showed no necrosis in the same region on day 3 any more. In the rats treated with 1 µg/100 g body weight/day of PTH and those treated with 3 µg/100 g body weight/day of PTH, bone resorption seemingly arose over wider range than in the rats treated with the vehicle. In the rats of these 2 groups, however, necrotic tissue was also observed.

10 Example 2

Experiment 1: Effect of continuous systemic PTH-infusion on experimental tooth movement

It has been already clarified that the continuous administration of PTH and the intermittent administration thereof differ from each other in their effect on bones. Thus, attempts were made to examine how PTH administered by either method would affect the experimental tooth movement. Use was made of 12 male Wister rats weighing 350 to 400 g. In the continuous PTH-administration group, an osmotic pump (2ML2, manufactured by Alzet, Palo Alto, Calif., USA) was subcutaneously implanted into the dorsal part of each rat followed by the continuous infusion of hPTH (1-34) (manufactured by Peptide Institute Inc., Mino) in a dose of 0.4 µg/100 g body weight/day or 4 µg/100 g body weight/day. As the control groups, use was made of 2 groups including (1) one to which the vehicle was continuously infused, and (2) an intermittent administration group to which 4 µg/100 g body weight/day of PTH was subcutaneously injected into the

dorsal part once a day. From the next day of the initiation of the PTH-administration, an ultra-elastic closed coil spring (509-21, manufactured by Tomy International, K.K., Tokyo) was put between the upper incisive tooth and the right first molar M_1 followed by the mesial traction of M_1 for 12 days (Fig. 6-A, B). After the initiation of the shifting, the precise impression of the upper jaw was taken every 3 days by using a silicone impression material (Exafine, manufactured by GC, Tokyo) and the distance between the first molar and the second molar (M_2) was measured on ultra-hard gypsum models (Fig. 6-C). After the completion of shifting over 12 days, arterial blood was collected from the abdominal aorta and various serum parameters were measured. After sacrificing each rat, the upper jaw and the right thickening bone were taken out. The upper jaw was decalcified and then cut into a paraffin section of 8 μm in thickness involving, in the direction of the major axis, the buccal mesial root and the buccal distal root followed by HE-staining and histological observation.

The bone mineral content and bone mineral density of the isolated femur were measured by dual-x-ray absorptometry (DCS-600, manufactured by Aloka, Tokyo, Japan).

Experiment 2: Effect of local administration of hPTH (1-34) in sustained release dosage form on experimental tooth movement

25 movement

To continuously release PTH having been locally administered, 0.1 $\mu\text{g}/\mu\text{l}$ and 1 $\mu\text{g}/\mu\text{l}$ PTH preparations containing 2% of methylcellulose as the base (PTH-MC) were

prepared by mixing respectively 0.2 $\mu\text{g}/\mu\text{l}$ and 2 $\mu\text{g}/\mu\text{l}$ PTH solutions in physiological saline with the same amount of 4% methylcellulose. Then these PTH-MC preparations were injected subperiosteally into the palatal mucosa in the mesial palatal side of M_1 in a dose of 1 μl every 2 days (corresponding respectively to 0.0125 $\mu\text{g}/100 \text{ g/day}$ and 0.125 $\mu\text{g}/100 \text{ g/day}$ as expressed in Experiment 1) with the use of microsyringes (manufactured by Hamilton) (Fig. 6-A). As the control groups, the following 3 groups were employed: (1) a group to which 1 μl of 2% methylcellulose (MC) alone was injected every 2 days into the same site; (2) one to which 1 μl of a 1 $\mu\text{g}/\mu\text{l}$ solution of PTH in physiological saline (PTH-physiological saline solution) was injected every 2 days into the same site; and (3) one to which 1 $\mu\text{g}/\mu\text{l}$ of PTH-MC was subcutaneously administered every 2 days to the dorsal part. The evaluation of the tooth movement and the histological observation were performed each in the same manner as the one employed in Experiment 1.

Results

- 20 1. Effect of continuous systemic PTH-infusion on experimental tooth movement (Figs. 7, 8 and 9).

On day 12 of the tooth movement, the control group showed a mesial shift of M_1 of $0.56 \pm 0.04 \text{ mm}$. In the PTH-administration groups, on the other hand, the shift of M_1 was promoted depending on the dose by the continuous infusion of PTH. Namely, the group with the continuous infusion of 4 $\mu\text{g}/100 \text{ g body weight/day}$ showed a shift of $1.01 \pm 0.09 \text{ mm}$, i.e., almost twice as much as that of the

control group.

2. Change with the passage of time in effect of continuous PTH infusion and intermittent PTH injection on experimental tooth movement (Fig. 10).

5 On day 3 of the tooth movement, the mesial shift of M_1 was slightly significantly promoted in both of the continuous PTH infusion group and the intermittent PTH injection group. After day 9 of tooth movement, this tooth shift-promoting effect was observed more remarkably in the 10 continuous PTH infusion group. In the intermittent PTH injection group, however, no significant difference from the control group was observed in the M_1 shifting distance after day 6.

15 3. Effect of local injection of sustained release PTH (1-34) on experimental tooth movement (Figs. 11, 12, 13 and 14).

On day 12 of tooth movement, the control group to which 2% methylcellulose alone had been given showed a mesial shift of M_1 of 0.54 ± 0.08 mm. In the groups with 20 the local administration of PTH-MC, on the other hand, the shift of M_1 was promoted depending on the concentration. Namely, the group to which 1 $\mu\text{g}/400$ g/day of PTH had been administered every 2 days showed a mesial shift of M_1 of 0.08 ± 0.11 mm, i.e., almost 1.6 times as much as that in 25 the control group. The group with the local administration of the PTH-physiological saline and the group with the subcutaneous administration of PTH-MC to the dorsal part each showed no promotion in tooth movement.

4. Change with the passage of time in effect of topical injection of sustained release PTH (1-34) on experimental tooth movement (Fig. 15).

When 1 µg/400 g of PTH (1-34) was locally injected
5 every 2 days, there was observed a tendency that the mesial M₁ shift was promoted since day 3 of the tooth movement. This tooth movement-promoting effect of PTH became more remarkable after day 9.

5. Histological findings in effects of the continuous PTH
10 (1-34) infusion and the local injection of sustained release PTH on orthodontic tooth movement (Figs. 16, 17 and 18)

In the control group, a vitrified denaturation was observed in the periodontal membrane part between the
15 alveolar septum and the distal root pressed by the traction force (Fig. 16-A). In the continuous infusion group, in contrast thereto, remarkable bone resorption was observed over a wide range in the distal side of the alveolar septum and no such necrotic tissue as observed in the control group
20 was found (Fig. 16-B). Compared with the control group, the continuous infusion group further showed energetic bone resorption in the mesial alveolar bone of M₁. On the other hand, the group with the local injection of sustained release PTH showed no such alveolar bone resorption over a
25 side range in compressed distal root side as observed in the group with the continuous systemic infusion (Fig. 18-B).

As described above, it has been proved that experimental tooth movement can be promoted by continuously

administering PTH systemically or by locally injecting a PTH preparation in a sustained release dosage form.

INDUSTRIAL APPLICABILITY

As described above, parathyroid hormone (PTH) or PTH derivatives have accelerated orthodontic tooth movement, which makes them useful as orthodontic remedies.

CLAIMS

1. A method for increasing tooth movement comprising administering to a subject in need thereof an effective amount of a parathyroid hormone or a derivative thereof.
2. The method as claimed in claim 1 wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 84.
3. The method as claimed in claim 2 wherein the parathyroid hormone is substantially pure parathyroid hormone.
4. The method as claimed in claim 2 wherein the parathyroid hormone is recombinant parathyroid hormone.
5. The method as claimed in claim 1 wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 34.
6. The method as claimed in claim 5 wherein the parathyroid hormone is substantially pure parathyroid hormone.
7. The method as claimed in claim 5 wherein the parathyroid hormone is recombinant parathyroid hormone.
8. A kit for increasing tooth movement comprising an effective amount of a parathyroid hormone or a derivative thereof and instructions for increasing tooth movement.
9. The kit as claimed in claim 8 wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 84.
10. The kit as claimed in claim 9 wherein the parathyroid hormone is substantially pure human parathyroid hormone.

- 00344662 0033633
11. The kit as claimed in claim 9 wherein the parathyroid hormone is recombinant parathyroid hormone.
 12. The kit as claimed in claim 8 wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 34.
 13. The kit as claimed in claim 12 wherein the parathyroid hormone is substantially pure parathyroid hormone.
 14. The kit as claimed in claim 12 wherein the parathyroid hormone is recombinant parathyroid hormone.
 15. In an orthodontic composition, the improvement comprising an effective amount of a parathyroid hormone or a derivative thereof for increasing tooth movement.
 16. The orthodontic composition as claimed in claim 15 wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 84.
 17. The orthodontic composition as claimed in claim 16 wherein the parathyroid hormone is substantially pure parathyroid hormone.
 18. The orthodontic composition as claimed in claim 16 wherein the parathyroid hormone is recombinant parathyroid hormone.
 19. The orthodontic composition as claimed in claim 15 wherein the parathyroid hormone is human parathyroid hormone comprising amino acids 1 to 34.
 20. The orthodontic composition as claimed in claim 19 wherein the parathyroid hormone is substantially pure parathyroid hormone.
 21. The orthodontic composition as claimed in claim 19

wherein the parathyroid hormone is recombinant parathyroid hormone.

ORTHODONTIC REMEDIES CONTAINING PTH

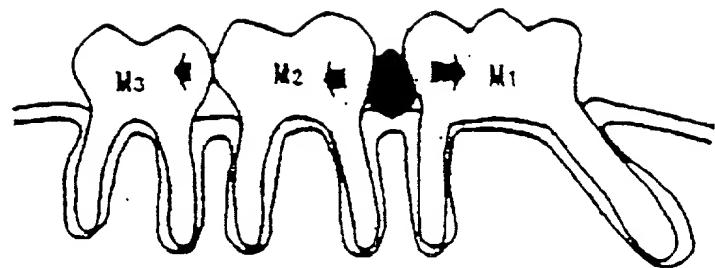
ABSTRACT

Orthodontic remedies containing parathyroid hormone (PTH) or one or more PTH derivatives as the active
5 ingredient(s).

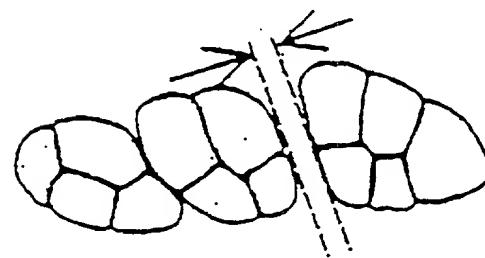
PCTC 282940

Fig. 1

(A)



(B)



contact gauge

(C)

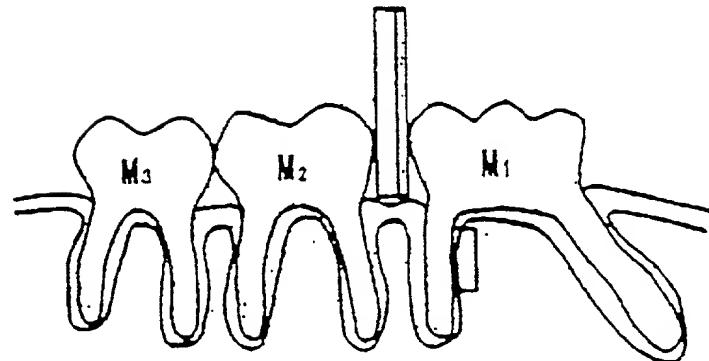


Fig. 2

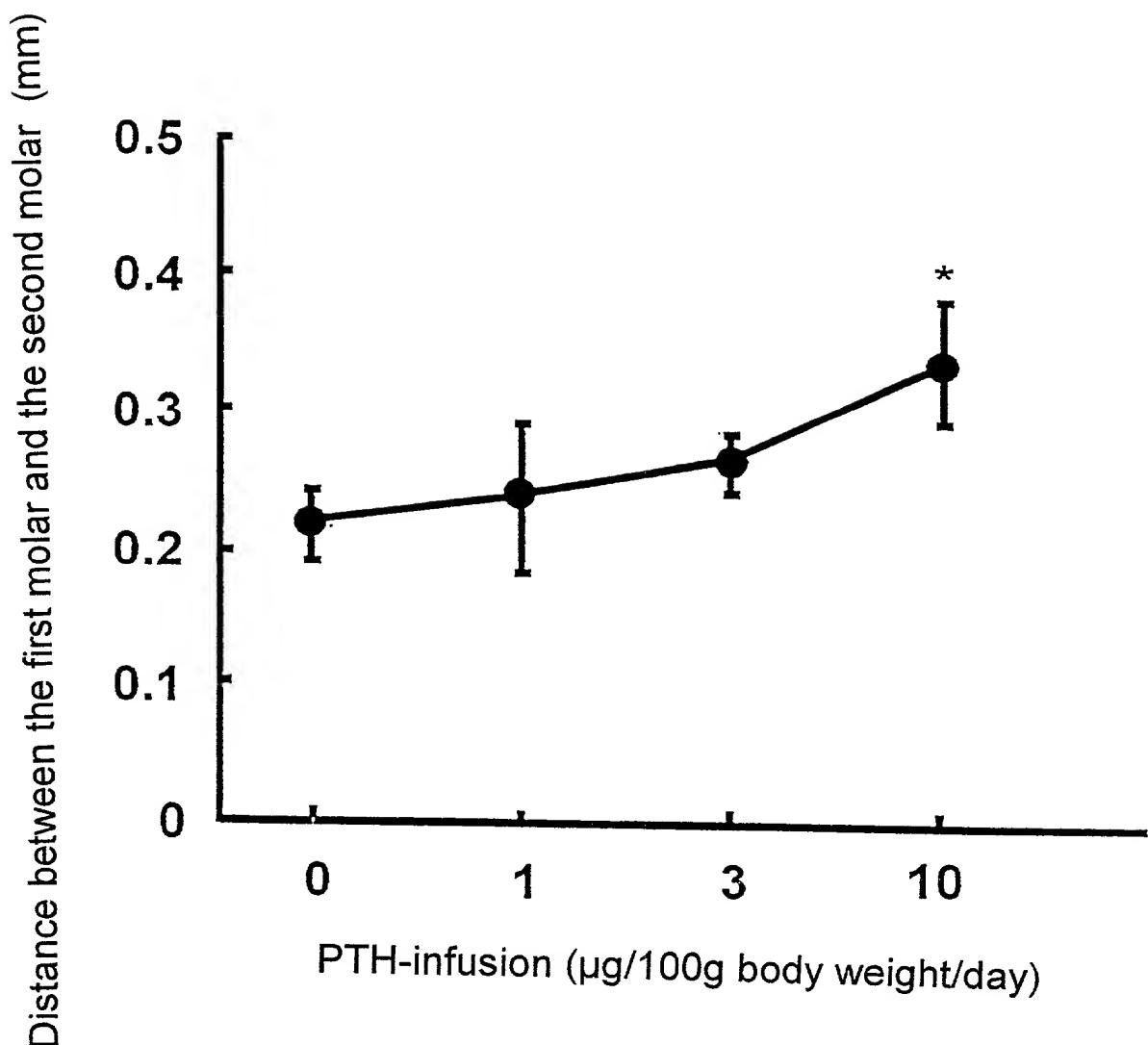


Fig. 3

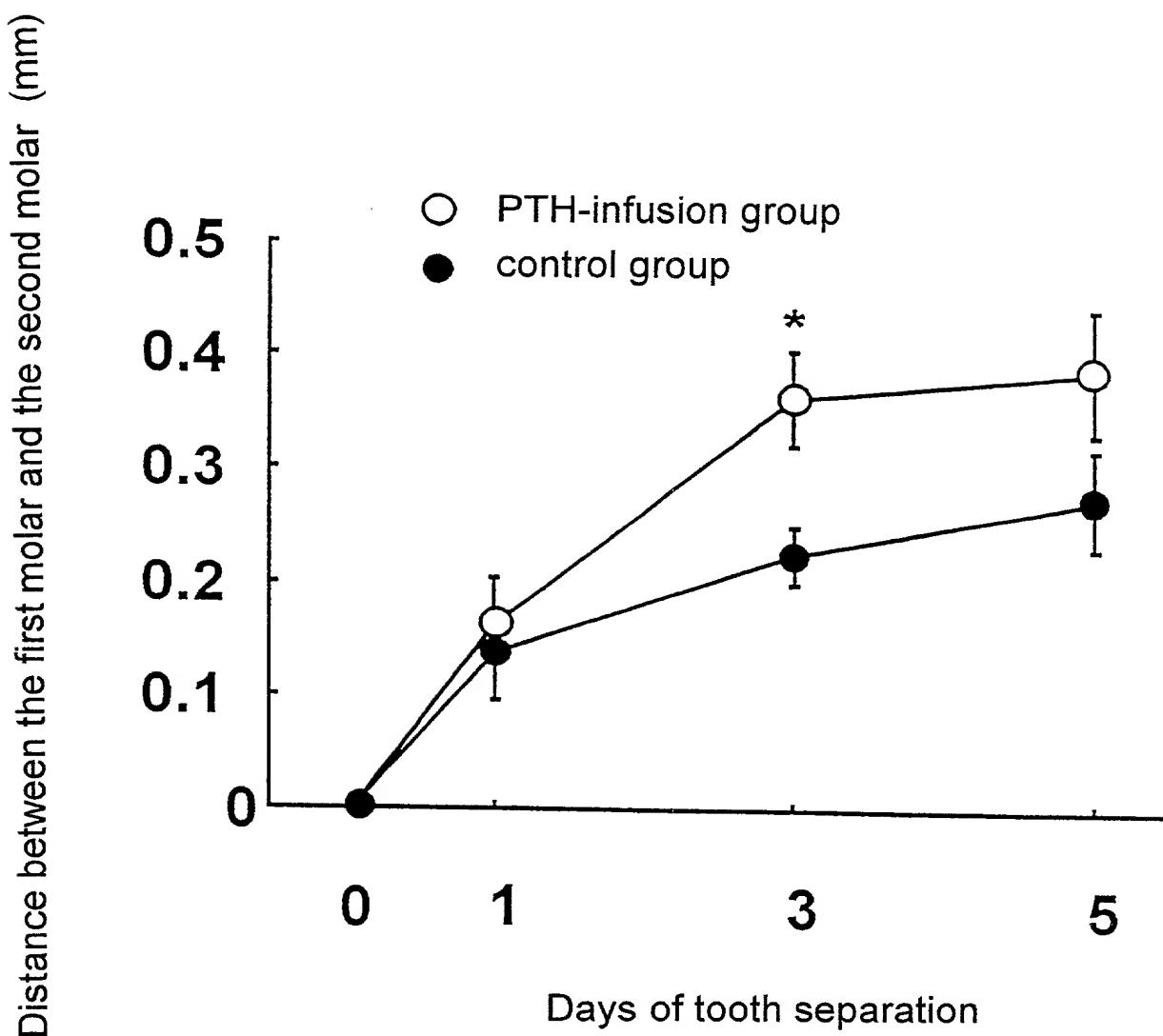


Fig. 4

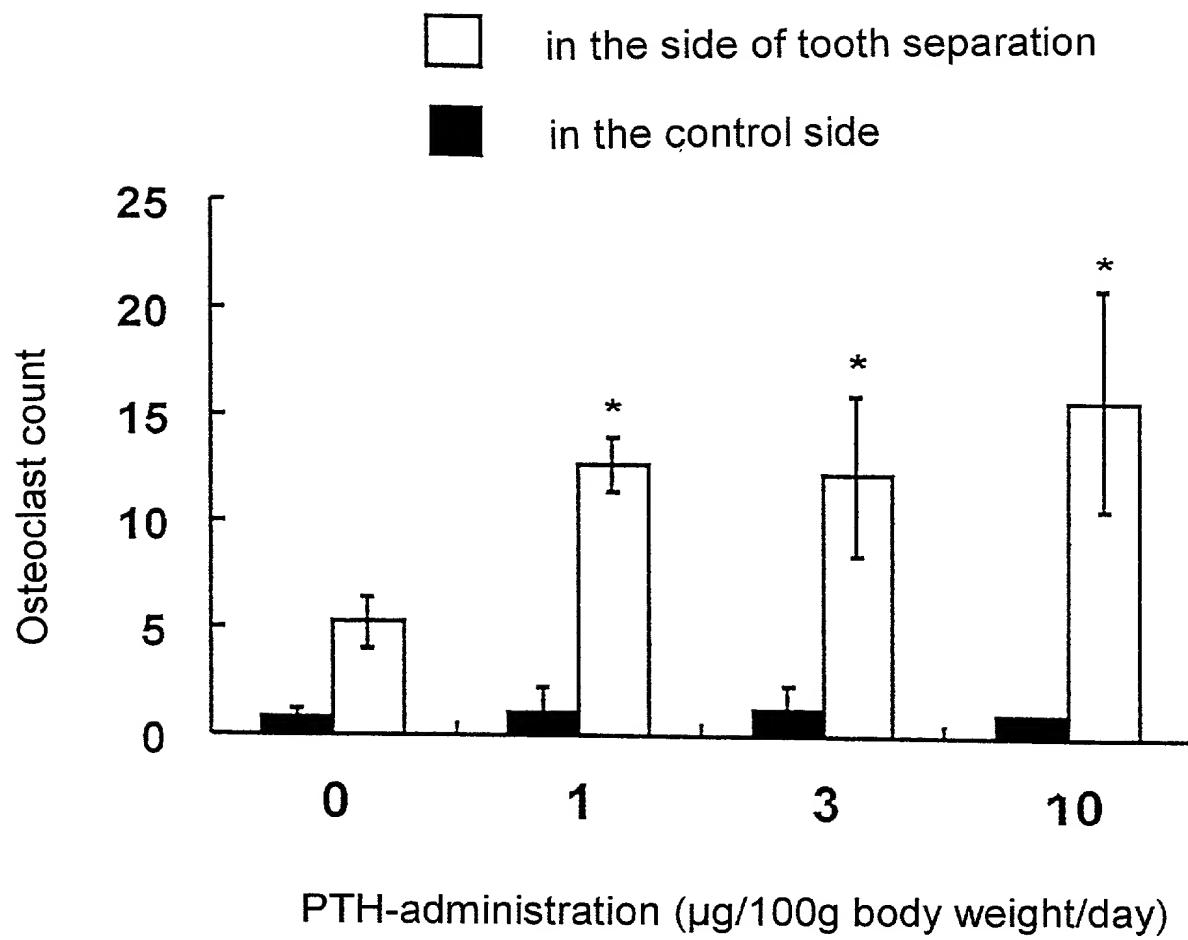


Fig. 5

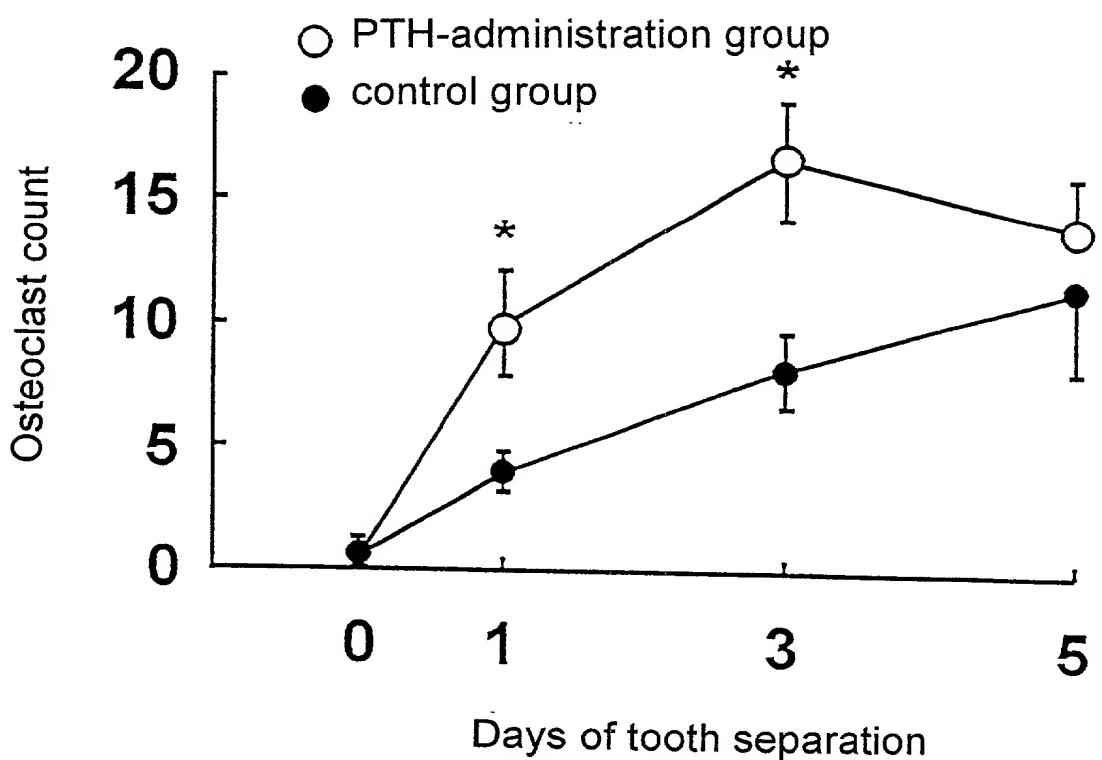
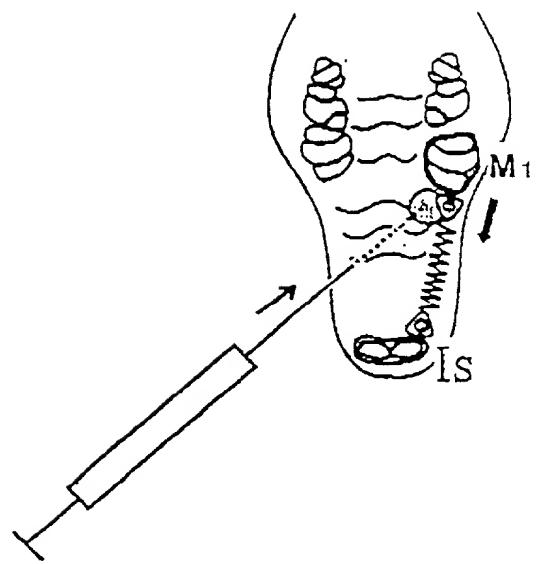


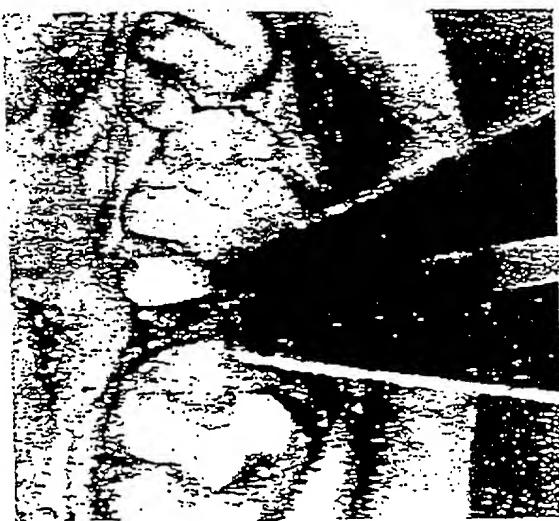
Fig. 6



(A)



(B)



(C)

Fig. 7

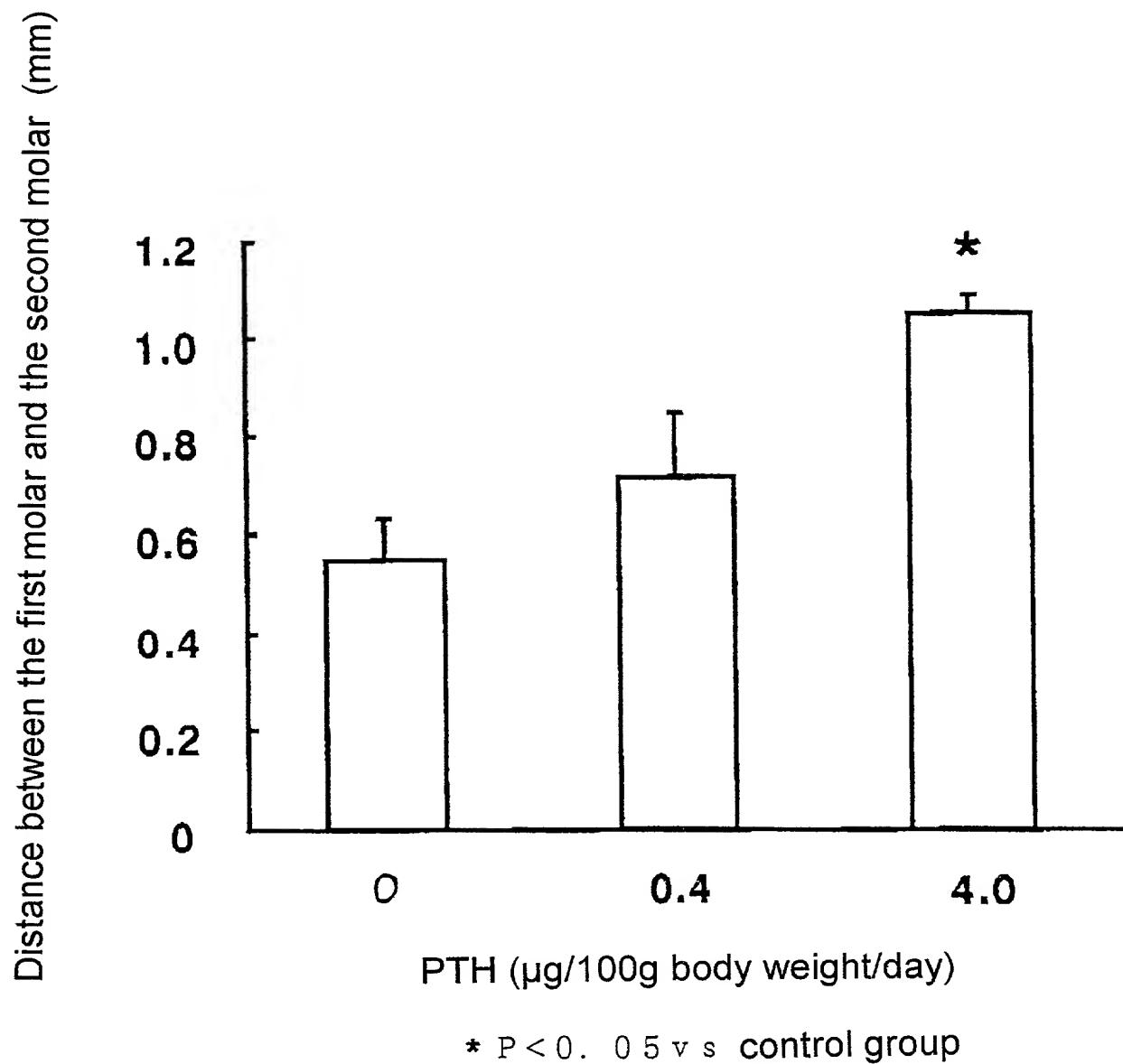


Fig. 8

(A) control group

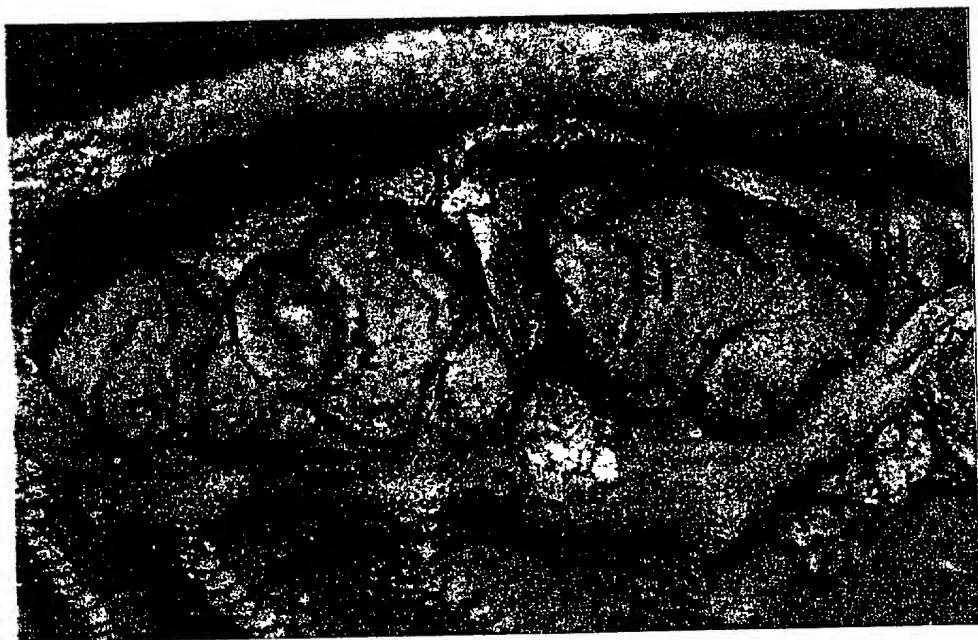


(B) PTH infusion group



Fig. 9

(C) PTH injection group



000000000000000000000000

Fig. 10

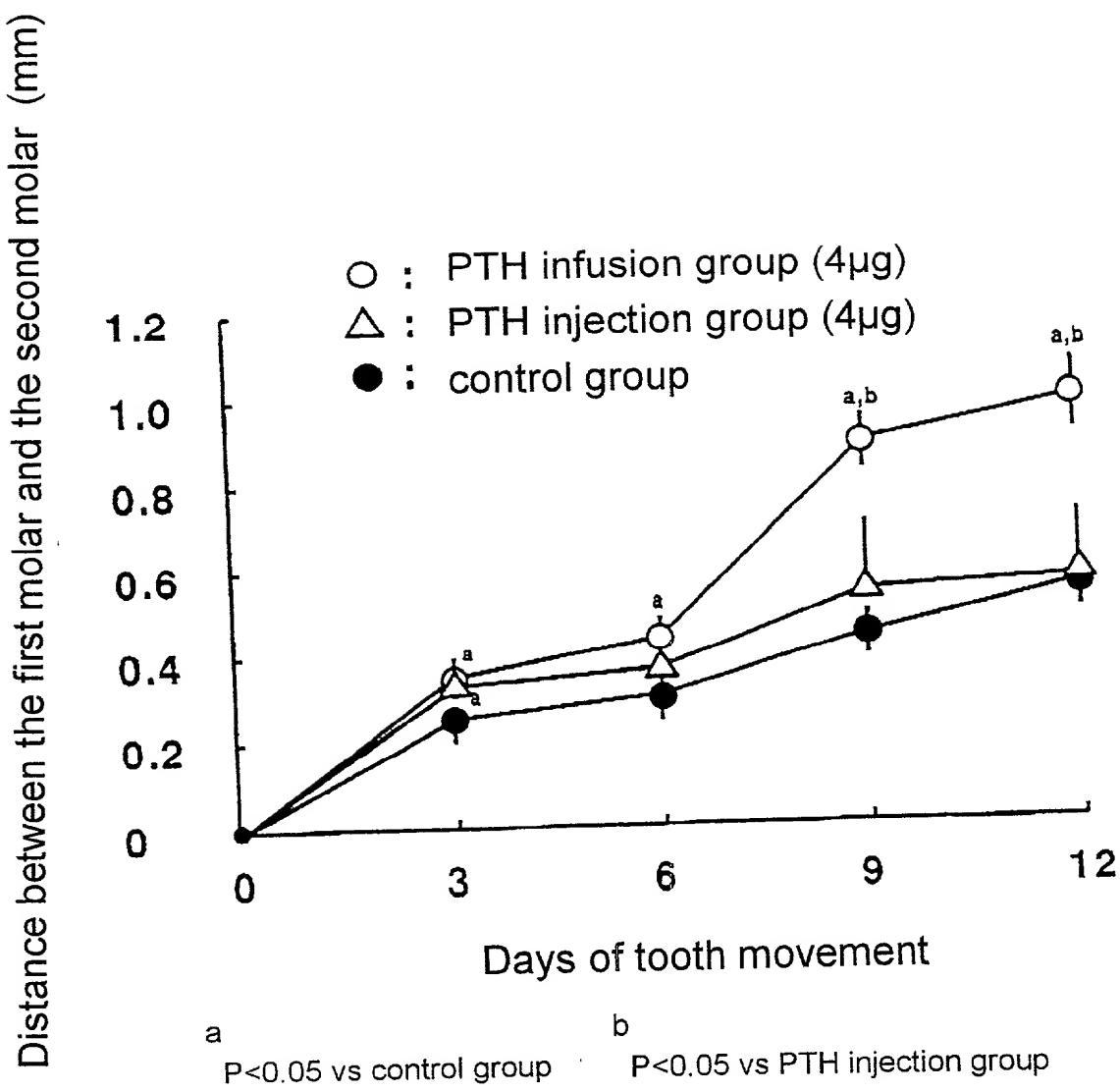


Fig. 11

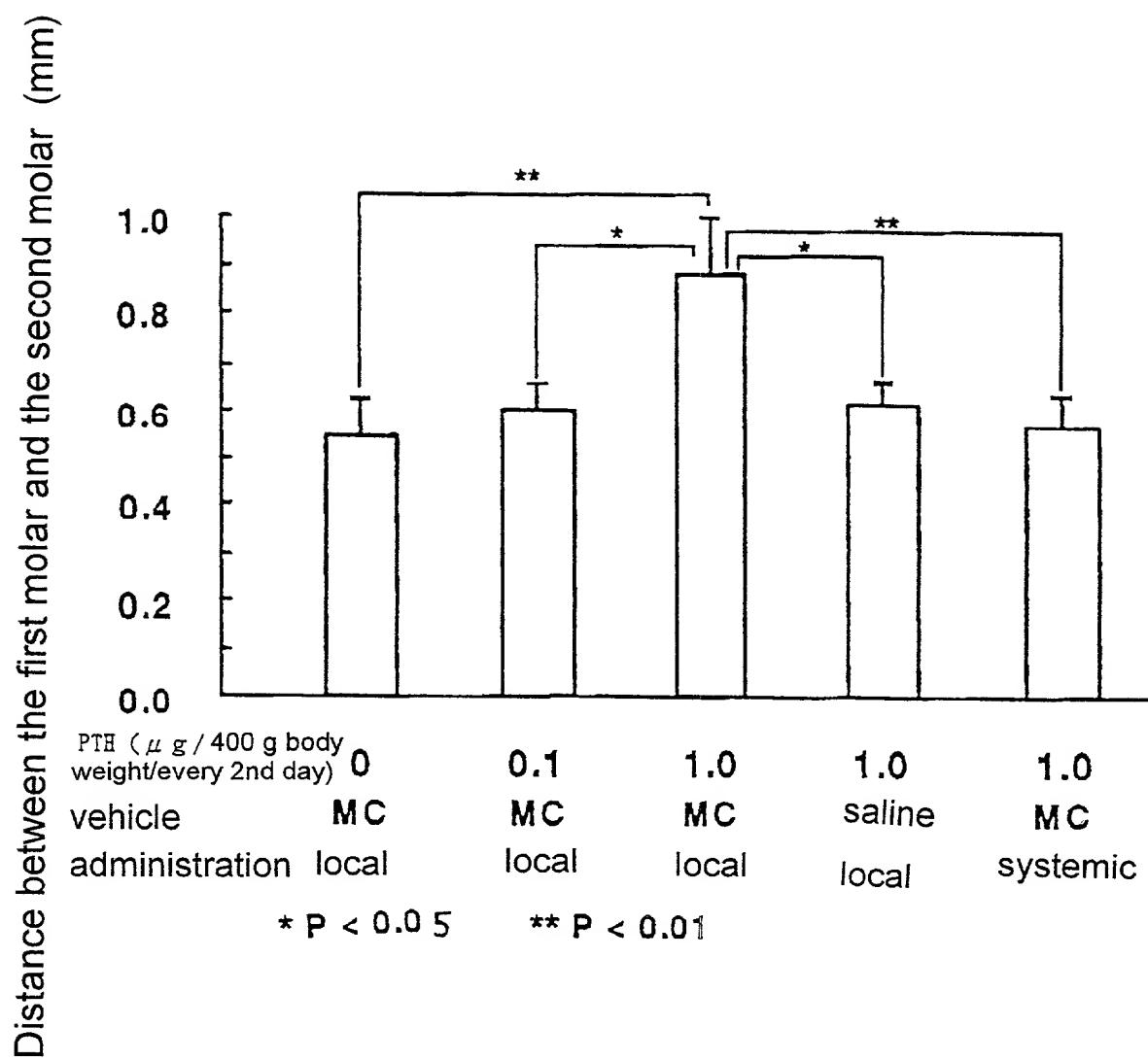


Fig. 12

(A)

PTH (μ g)	vehicle	injection
0	2% MC	local



(B)

PTH (μ g)	vehicle	injection
0.1	2% MC	local

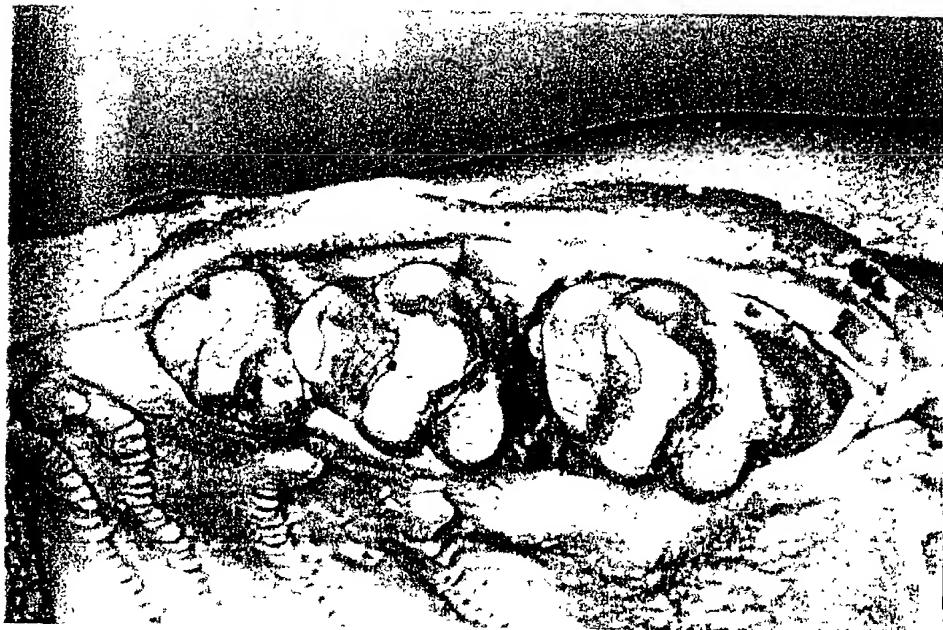
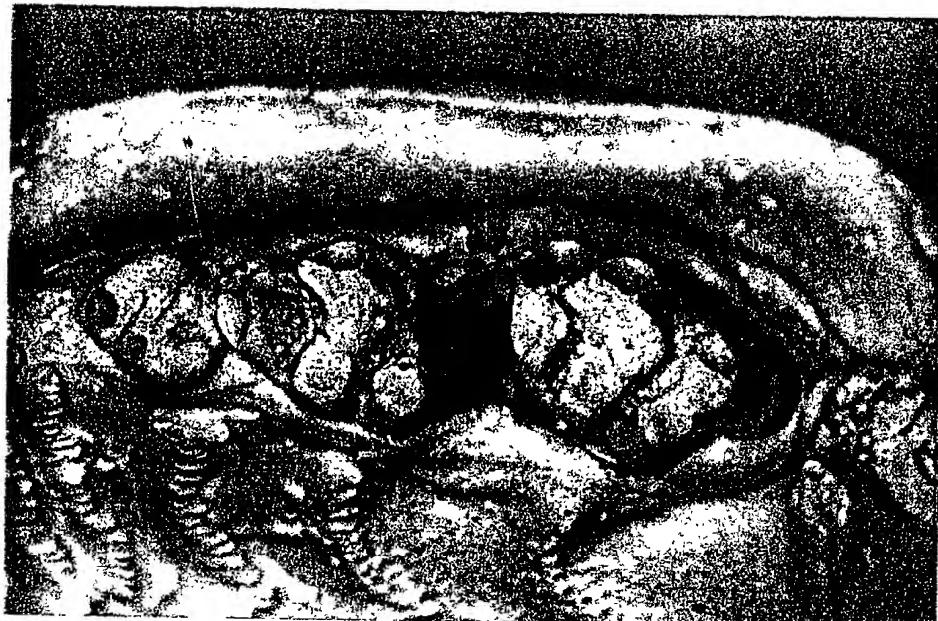


Fig. 13

(A)

PTH (μ g): vehicle : injection		
1.0	2% MC	local



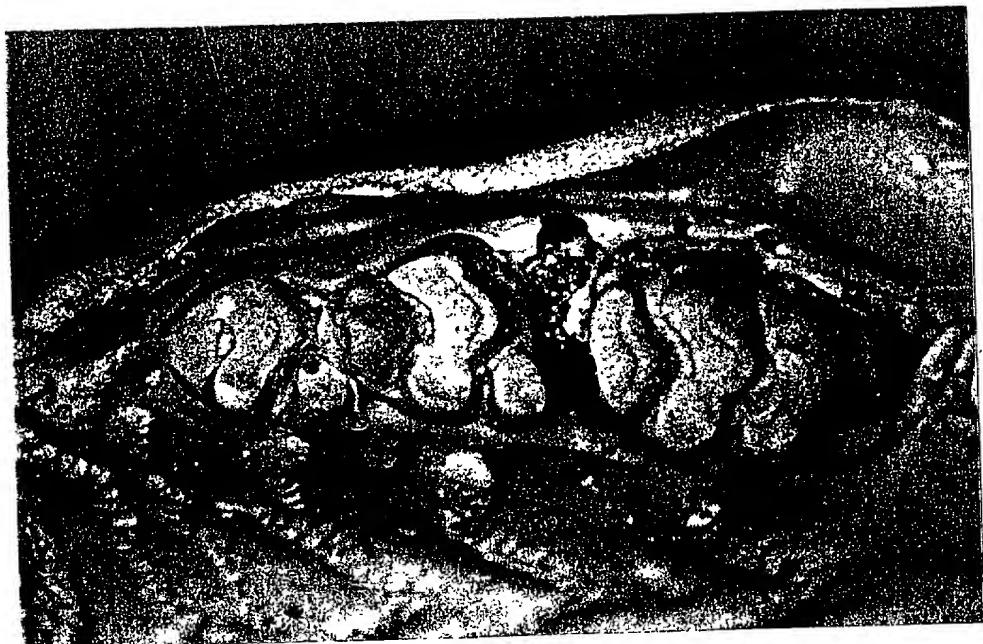
(B)

PTH (μ g): vehicle : injection		
1.0	saline	local



Fig. 14

PTH (μ g)	vehicle	injection
1.0	2% MC	systemic



009344288 "062598

Fig. 15

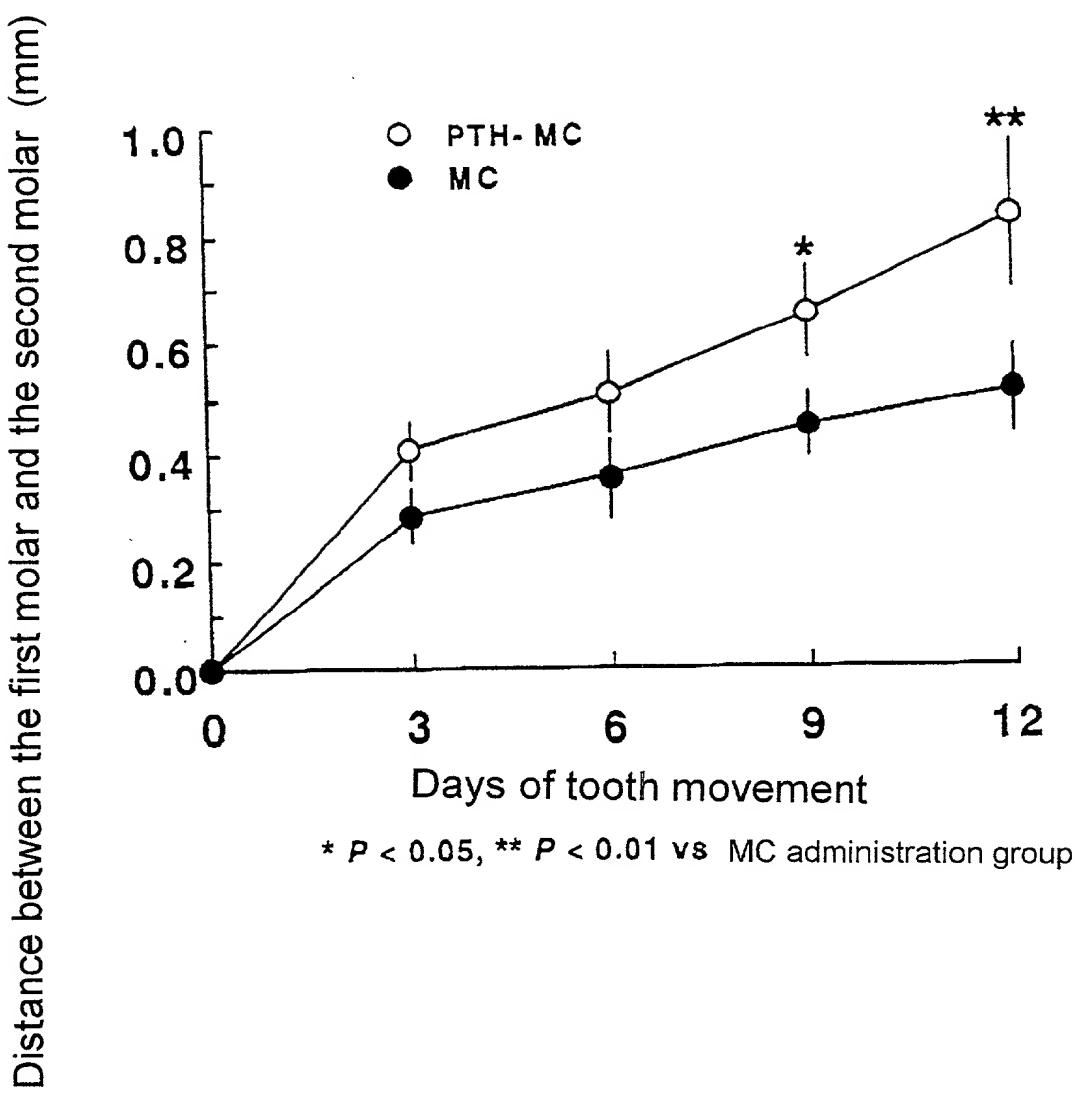


Fig. 16

< A > control group



< B > PTH infusion group



Fig. 17

< C > PTH injection group

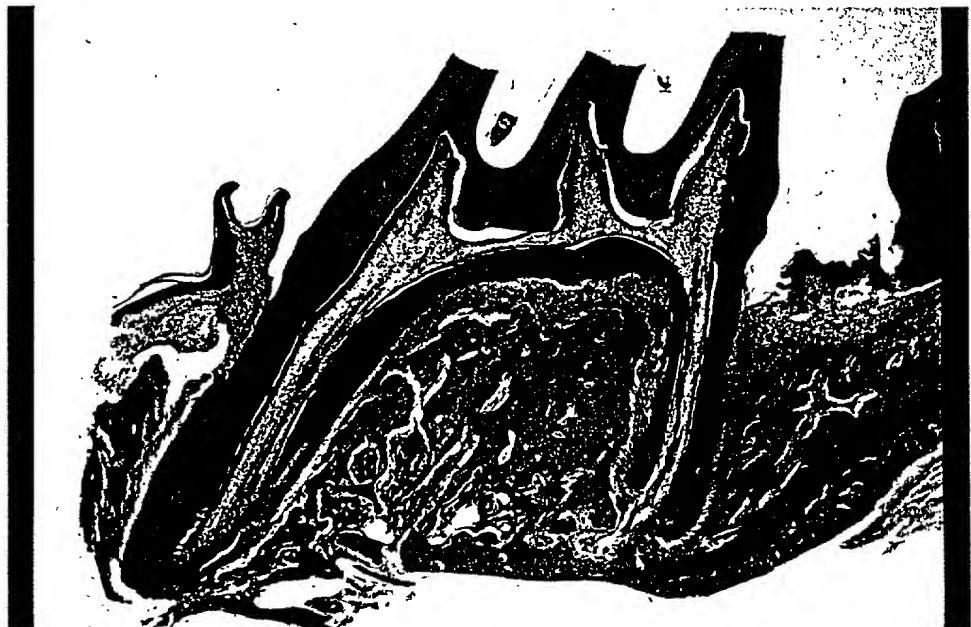


Fig. 18

local injection

(A)

2% MC

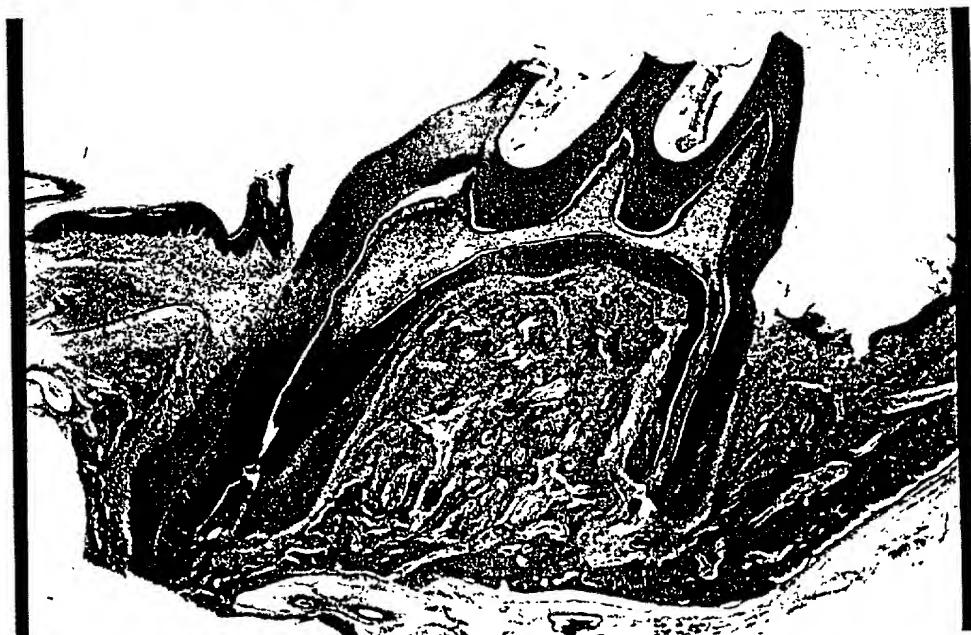


(B)

PTH

+

2% MC



Declaration and Power of Attorney For Patent Application

特許出願宣言書及び委任状

Japanese Language Declaration

日本語宣言書

下記の氏名の発明者として、私は以下の通り宣言します。 As a below named inventor, I hereby declare that:

私の住所、私書箱、国籍は下記の私の氏名の後に記載された通りです。 My residence, post office address and citizenship are as stated next to my name.

下記の名称の発明に関して請求範囲に記載され、特許出願している発明内容について、私が最初かつ唯一の発明者（下記の氏名が一つの場合）もしくは最初かつ共同発明者であると（下記の名称が複数の場合）信じています。

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

ORTHODONTIC REMEDIES CONTAINING

PTH

上記発明の明細書（下記の欄で x 印がついていない場合は、the specification of which is attached hereto unless the following 本書に添付）は、

____月____日に提出され、米国出願番号または特許協定条約
国際出願番号を_____とし、
(該当する場合) _____に訂正されました。

was filed on _____
as United States Application Number or
PCT International Application Number
_____ and was amended on
_____ (if applicable).

私は、特許請求範囲を含む上記訂正後の明細書を検討し、
内容を理解していることをここに表明します。

I hereby state that I have reviewed and understand the contents
of the above identified specification, including the claims, as
amended by any amendment referred to above.

私は、連邦規則法典第37編第1条56項に定義されると
おり、特許資格の有無について重要な情報を開示する義務が
あることを認めます。

I acknowledge the duty to disclose information which is material
to patentability as defined in Title 37, Code of Federal
Regulations, Section 1.56.

Japanese Language Declaration (日本語宣言書)

私は、米国法典第35編119条(a)-(d)項又は365条(b)項に基き下記の、米国以外の国の少なくとも一ヵ国を指定している特許協力条約365(a)項に基づく国際出願、又は外国での特許出願もしくは発明者証の出願について外国優先権をここに主張するとともに、優先権を主張している、本出願の前に出願された特許または発明者証の外国出願を以下に、枠内をマークすることで、示しています。

Prior Foreign Application(s)

外国での先行出願

Priority Claimed 優先権主張			
<input checked="" type="checkbox"/> Yes はい	<input type="checkbox"/> No いいえ	<input type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> Yes はい	<input type="checkbox"/> No いいえ	<input type="checkbox"/>	<input type="checkbox"/>

私は、第35編米国法典第119条(e)項に基いて下記の米国特許出願規定に記載された権利をここに主張いたします。

(Application No.) (出願番号)	(Filing Date) (出願日)	(Application No.) (出願番号)	(Filing Date) (出願日)
-----------------------------	------------------------	-----------------------------	------------------------

私は、下記の米国法典第35編120条に基いて下記の米国特許出願に記載された権利、又は米国を指定している特許協力条約365条(c)に基づく権利をここに主張します。また、本出願の各請求範囲の内容が米国法典第35編112条第1項又は特許協力条約で規定された方法で先行する米国特許出願に開示されていない限り、その先行米国出願書提出日以降で本出願書の日本国内または特許協力条約国提出日までの期間中に入手された、連邦規則法典第37編1条56項で定義された特許資格の有無に関する重要な情報について開示義務があることを認識しています。

PCT/JP97/04891	26/12/1997	Pending
(Application No.) (出願番号)	(Filing Date) (出願日)	(Status: Patented, Pending, Abandoned) (現況: 特許許可済、係属中、放棄済)
(Application No.) (出願番号)	(Filing Date) (出願日)	(Status: Patented, Pending, Abandoned) (現況: 特許許可済、係属中、放棄済)

私は、私自身の知識に基づいて本宣言書中で私が行なう表明が真実であり、かつ私の入手した情報と私の信じるところに基づく表明が全て真実であると信じていること、さらに故意になされた虚偽の表明及びそれと同等の行為は米国法典第18編第1001条に基づき、罰金または拘禁、もしくはその両方により処罰されること、そしてそのような故意による虚偽の声明を行なえば、出願した、又は既に許可された特許の有効性が失われることを認識し、よってここに上記のごとく宣言を致します。

I hereby claim foreign priority under Title 35, United States Code, Section 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

I hereby claim the benefit under Title 35, United States Code, Section 119(e) of any United States provisional application(s) listed below.

I hereby claim the benefit under Title 35, United States Code, Section 120 of any United States application(s), or 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code Section 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of application.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Japanese Language Declaration (日本語宣言書)

委任状： 私は下記の発明者として、本出願に関する一切の手続きを米特許商標局に対して遂行する弁理士または代理人として、下記の者を指名いたします。（弁護士、または代理人の氏名及び登録番号を明記のこと）

Stephen A. Bent, Reg. No. 29,768
 David A. Blumenthal, Reg. No. 26,257
 William T. Ellis, Reg. No. 26,874
 John J. Feldhaus, Reg. No. 28,822
 Patricia D. Granados, Reg. No. 33,683
 John P. Isaacson, Reg. No. 33,715
 Eugene M. Lee, Reg. No. 32,039
 Richard Linn, Reg. No. 25,144
 Peter G. Mack, Reg. No. 26,001

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith (*list name and registration number*).

Brian J. McNamara, Reg. No. 32,789
 Sybil Meloy, Reg. No. 22,749
 George E. Quillin, Reg. No. 32,792
 Colin G. Sandercock, Reg. No. 31,298
 Bernhard D. Saxe, Reg. No. 28,665
 Charles F. Schill, Reg. No. 27,590
 Richard L. Schwaab, Reg. No. 25,479
 Arthur Schwartz, Reg. No. 22,115
 Harold C. Wegner, Reg. No. 25,258

書類送付先

Send Correspondence to:

Foley & Lardner
 3000 K Street, N.W.
 P.O. Box 25696
 Washington, DC 20007-8696

Foley & Lardner
 3000 K Street, N.W.
 P.O. Box 25696
 Washington, DC 20007-8696

直接電話連絡先：（名前及び電話番号）

Direct Telephone Calls to: (*name and telephone number*)

(202)672-5300

(202)672-5300

唯一または第一発明者名		Full name of sole or first inventor Shunichi SOMA	
発明者の署名	日付	Inventor's signature <i>Shunichi Soma</i>	Date June 18, 1999
住所	Residence Osaka, Japan		
国籍	Citizenship Japanese		
私書箱	Post Office Address 2-22-16-403, Tsukamoto,		
	Yodogawa-ku, Osaka-shi, Osaka 532-0026 Japan		
第二共同発明者	Full name of second joint inventor, if any Masahiro IWAMOTO		
第二共同発明者の署名	日付	Second inventor's signature <i>Masahiro Iwamoto</i>	Date June 18, 1999
住所	Residence Osaka, Japan		
国籍	Citizenship Japanese		
私書箱	Post Office Address 4-6-10-606, Aoshinke,		
	Minoo-shi, Osaka 562-0024 Japan		

(第三以降の共同発明者についても同様に記載し、署名をすること)
 (Supply similar information and signature for third and subsequent joint inventors.)

Japanese Language Declaration

第三共同発明者氏名	Full name of third joint inventor Kojiro KURISU		
同発明者の署名	日付	Inventor's signature	Date June 18 1999
住所	Residence Nara-ken, Japan		
国籍	Citizenship Japanese		
郵便の宛先	Post Office Address 2-1-7, Mayumiminami, Ikoma-shi, Nara 630-0123 Japan		
第四共同発明者	Full name of fourth joint inventor, if any Yoshinobu HIGUCHI		
同発明者の署名	日付	Inventor's signature	Date June 18 1999
住所	Residence Shizuoka-ken, Japan		
国籍	Citizenship Japanese		
郵便の宛先	Post Office Address c/o Chugai Seiyaku Kabushiki Kaisha of 135, Komakado 1-chome, Gotenba-shi, Shizuoka 412-0038 Japan		

第五共同発明者氏名	Full name of fifth joint inventor		
同発明者の署名	日付	Inventor's signature	Date
住所	Residence		
国籍	Citizenship		
郵便の宛先	Post Office Address		
第六共同発明者	Full name of sixth joint inventor, if any		
同発明者の署名	日付	Inventor's signature	Date
住所	Residence		
国籍	Citizenship		
郵便の宛先	Post Office Address		